

Production of nickel oxide nanoparticles by laser and study of antibacterial properties against wound infection

Sara F. Abbas ^{a *}, Adawiya J. Haider ^b, Sharafaldin Al-Musawi ^c, Murtadha K. Selman ^d, Jha Prafulla K ^e, and Ali A. Al-Muntaser ^f

^aLaser and Optoelectronics Engineering Department, University of Technology- Iraq

^bApplied Science Department/Laser Science and Technology Branch, University of Technology- Iraq

^cCollege of Food Sciences, Al-Qasim Green University, Babylon Province, Iraq

^dMinistry of Education - Babylon Education Directorate, Babylon Province, Iraq

^eDepartment of Physics, Faculty of Science, M. S. University of Baroda, India

^fPhysics Department, Sana'a University, Yemen

* Corresponding author. Tel.: +964-772-368-7684; e-mail: sarafadhel1989@gmail.com

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ABSTRACT

Nanostructure materials have a variety of technical applications that come from their remarkable size-dependent properties as compared to particle range size. Nickel oxide (NiO) nanoparticles (NPs) acquire huge attention related to their promise usage in many fields. NiO NPs are produced using pulsed laser ablation (PLA) in distilled water (DW), a highly controlled and eco-friendly method. The manufactured NiO NPs were investigated by various techniques to study their physio-chemical properties. Structural analysis of NiO nanoparticles (NiO NPs) using X-ray diffraction (XRD) revealed that they possess a face-centered cubic (FCC) crystal structure. The formation of NiO NPs is indicated by Fourier transform infrared spectroscopy (FTIR) which presents a clear FTIR peak at 688 cm⁻¹. Field Emission Scanning Electron Microscope (FESEM) displays the common morphology with spheroids-like particles coalesced layer structure, and the mean particle size is 44 nm. Tauck's formula was employed to estimate the energy band gap, which was found to be approximately 5.2 eV. Dynamic light scattering (DLS) shows that the particle size was 38.9 nm and that the NiO NPs had good stability. The zeta potential was negatively charged at 12.5 mV. *Staphylococcus aureus* (*S. aureus*) and *Escherichia coli* (*E. coli*) were examined for their antibacterial efficacy against the NiO NPs. The current investigation demonstrates strong antibacterial activity that may be investigated in the next clinical interventions.

Keywords: Nanotechnology, Antibacterial, Nickel oxide, Laser ablation in liquid

1. INTRODUCTION

Recently, nanotechnology has become the focus of scientists' attention due to its numerous technical applications in various fields [1–3]. Nanomaterials have incredibly interesting and practical qualities that come from their unique optical, electronic, and chemical properties [4], [5]. NPs have exhibited substantial antibacterial action, making them one of the most promising novel antimicrobial agents.

Alarming rates of antibiotic resistance are a result of the inappropriate and extensive use of antibiotics for both preventative and therapeutic purposes [6, 7]. Scientists are searching for novel antimicrobial medications as a result of developing antibiotic resistance of the common bacterial strains [8–10]. As a result, new antibacterial techniques must be developed to combat antibiotic-resistant microorganisms. Metals oxide nanoparticles (MeO NPs) are thought to be superior antibacterial medications because of their great stability, durability, and reduced toxicity to mammalian cells [11]. Various metals and metal oxides, like zinc, silver, copper, nickel, magnesium, titanium, etc., have ascended attention in environmental purification, food

packaging, healthcare and biological and medical fields due to their ability to kill both bacterial strains [12, 13].

NiO is categorized as a p-type semiconductor with a wide energy gap ranging from 3.6 to 4.0 eV. NiO is used extensively in many different chemical and physical applications [14, 15]. These applications include gas sensors, adsorbents, electronics, catalytic materials, and magnetic and antibacterial agents [16].

Recent research concentrated on studying NiO NPs for their special qualities. NiO NPs exhibit encouraging promise in different biomedical applications. Research is being conducted to explore the antibacterial features of NiO NPs, along with their potential applications in cancer diagnosis and treatment. The antibacterial action of NiO NPs was studied by Ramasami *et al.* [17] against fungal (*C. albicans*) and (*B. cereus* and *K. aerogenes*) bacterial strains. Talebian *et al.* [18] investigated the antibacterial action of NiO NPs versus food-borne *S. aureus* and *E. coli* bacteria. Vijaya Kumar *et al.* evaluated the antibacterial efficacy of NiO NPs versus a wider variety of harmful microorganisms [17]. Isa Khan *et al.* [19] used NiO NPs to demonstrate how sensitive bacteria are to various bacterial strains (such as *S. aureus*

and *E. coli*). Using the agar diffusion method, Helan *et al.* [20] verified the cytotoxic effect of NiO NPs towards pathogenic bacteria including *S. aureus* and *E. coli*. NiO NPs differ greatly from bulk concerning their physio-chemical characteristics because of their quantum size and surface effects [21]. The compounds, including NiO NPs, have been applied extensively in several investigations, and their characteristics and uses have been thoroughly documented up to this point [22, 23]. Specifically, NiO NPs have generated an interest in biological and medical sciences due to their great biological properties, including their low cytotoxic impact. Because of their distinctive characteristics, they are more appropriate for applications where common materials have high toxicity or ineffectiveness. Nevertheless, there is a dearth of thorough research on the antibacterial qualities of NiO and its nanostructure. Understanding the safety and effectiveness of NiO NPs more in detail can make them important tools for the future development of nanomedicine [23].

The mechanism of antibacterial activity of NiO NPs is not recognized [24]. The antibacterial features of NiO NPs are mediated by several methods. Reactive oxygen species (ROS) production upon contact with bacterial cells is one of the most important mechanisms. These ROS cause oxidative stress, which harms lipids, proteins, and DNA in cells and eventually kills the bacteria [25, 26]. The heightened antibacterial activity of NiO NPs is partly owed to their unique nanostructure characteristics, which contain high surface area-per-volume proportion and small size. All these collective actions render NiO NPs highly potent against bacteria inclusive of antibiotic-resistant strains and biofilm-associated bacteria. It also minimizes the chances that encountered bacteria will develop resistance to a particular target; thus, NiO NPs may find useful applications

in the fields of medicine and other applications such as in the development of medical equipment and materials, wound healing dressings, and coatings. These characteristics let the nanoparticles engage with bacterial cells more effectively, which raises their effectiveness [27].

Many methods are used to produce NiO NPs, such as sol-gel, microwave irradiation, solid-state reaction, thermal decomposition, laser ablation, etc. [28–30]. Of all the mentioned synthesis processes, the PLA method has many advantages. In PLA, strategies for regulating pressure or heat are not necessary [31–33]. PLAL is a versatile method for producing many types of NPs, including semiconductors, alloys, oxides, and noble metals [34–36].

This technique involves developing a plasma plume that is fully immersed in a liquid medium while utilizing high-intensity PLA to establish an interaction between laser matter and the target's surface [37, 38]. The PLA method and plasma produced are shown in Figure 1(c).

This study aimed to utilize a straightforward method to produce NiO NPs as an effective antibacterial agent and examine their impact on common wound-associated pathogenic bacteria. The research showcases the fabrication of NiO NPs using the PLA method, presenting results from structural, optical, chemical, and zeta potential characterization analyses. Additionally, it offers insights into the antibacterial effectiveness of NiO NPs against the pathogenic strains associated with wound infections. This comprehensive investigation details the various properties of NiO NPs and emphasizes their potential for diverse biological applications, particularly in wound care and antibacterial treatments.

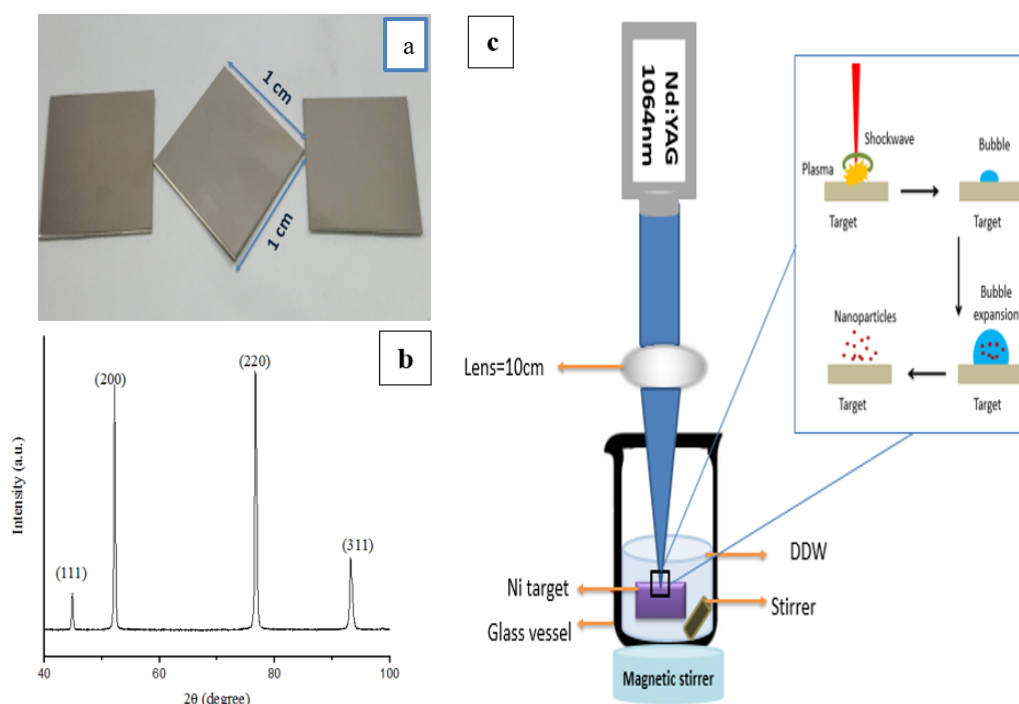


Figure 1. (a) Nickel sheet after cutting. (b) XRD analysis of the nickel sheet. (c) Schematic of PLA process

2. SAMPLE PREPARATION METHOD

A high purity (99.999%) nickel sheet was supplied from (BDH Chemical Ltd pool, England). The XRD pattern of the nickel target is shown in Figure 1(b). The PLA method was employed in this work to produce nanostructured NiO. The metal sheet was cut into small parts (1x1) cm, as seen in Figure 1(a). The nickel target was cleaned by using methanol and DW. The target was put inside a glass container that had (5 ml) of DW. In this method, a Q-switched Nd:YAG laser (Huafei Tongda Technology, DIAMOND-288 model EPLS) with a wavelength of 1064 nm, pulse duration of 10 nanoseconds, and repetition rate of 1 Hz was used to generate the nanostructure. A convex lens with a 10 cm focal length was employed to focus 32 J/cm² laser fluence for every pulse onto the target while mechanical stirring was applied during the process. Five experiments were repeated for each sample, so the results were similar.

3. CHARACTERIZATION TECHNIQUES

Different methods were applied to study the properties of the produced NiO NPs. The XRD pattern of the NiO NPs was analyzed using an AERIS PAN alytical XRD instrument with a Cu-K α radiation source, over a 2 θ range of (10–80) degrees. FTIR spectroscopy using the 8000 Series Shimadzu was employed to detect bond vibrations of the NPs in the 400–4000 cm⁻¹ range. A double-beam UV VIS spectrophotometer (Shimadzu UV-1800) was used to record the the optical absorption spectrum, and the energy band gap was determined using Tuac's formula. The nanostructures' morphology was examined using an Axia ChemiSEM SEM. EDX was used to verify the chemical construction of the prepared sample. DLS (Malvern Zetasizer ZS, Malvern, UK) was used to determine the particle diameter and zeta potential.

4. ANTIBACTERIAL ASSAY

The agar well diffusion assay was employed to study the antibacterial qualities of the NiO NPs and determine their characteristics. It was based on Selecting these bacteria, namely *S. aureus* and *E. coli*, which belong to gram-positive and gram-negative groups, respectively. The amount of NiO NP solution (25, 50, 100, 150, and 200 μ g/ml) was diluted in the experimental plates. After 24h of enculture in sterile Petri dishes at 37° C for each growth plate, the zone of inhibition was checked. By 24 h, the efficacy of the concentration was calculated, and the diameter of the growth inhibition was measured as encircling zones. Finally, the distance between the zone (mm) around each well was measured.

5. RESULTS AND DISCUSSIONS

The surface of Ni interacts with PLA to produce a plasma plume that splashes into liquid media. The alteration of water colour can provide an initial indication of the formation of NiO NPs. An intense shift in the absorption

spectra of NiO NPs is observed around 250–300 nm, as shown in Figure 2. Determining the energy band gap is essential, as it provides insight into a material's electronic, optical, and thermal properties. The optical band gap of NiO is broader than that of the bulk (3.65 eV). This may relate to the quantum confinement phenomena; the peak absorption wavelength can change based on the size of the NPs. The absorption spectra change when the size of the NPs reduces because the electrons' energy levels become quantized. When it comes to absorption peaks, smaller NPs typically exhibit a bluer shift than larger ones. This effect results from the creation of new energy levels that increase the value of band gap energy because of gaps in the intergranular areas or chemical flaws [39]. The optical band gap was estimated using the standard Tauc's relation [40]:

$$(\alpha h\nu) = A (h\nu - E_g)^n \quad (1)$$

where "A" is a constant, and "h ν " is the incident photon energy, n equals ½ for direct transition. The intercept point at = 0 can be used to calculate the E_g value from the Plotting h ν on the x-axis and $(\alpha h\nu)^2$ on the y-axis, presented in Figure 2 (inset). The E_g value of 5.2 eV that NiO NPs had is consistent with the findings of other writers [17, 41].

FTIR was performed on the materials to determine their chemical bonds and functional groups. Figure 3 illustrates the FTIR spectra of NiO NPs as a synthesis that shows some significant absorption peaks. The peaks observed in Figure 3 are at 3435, 2071, 1634, and 688 cm⁻¹. The peak at 3435 cm⁻¹ corresponds to the stretching vibrations of H₂O molecules, while the peak at 1634 cm⁻¹ is associated with their bending mode [42, 43]. The investigation showed that the presence of carbon dioxide may have come from atmospheric contamination at 2071 cm⁻¹.

The peak at 688 cm⁻¹ ascribes to NiO NPs Ni–O stretching vibration mode [44, 45]. Table 1 shows the FTIR information. Since the absorption band is so wide, the NiO NPs may be quite small and crystalline [46]. Because the particles studied in this work were much smaller than bulk NiO, the NiO NPs' IR peak of the Ni–O stretching vibration was moved near the blue region. This concept is relayed to the reduced size of particles, quantum confinement effects, or modifications to the surface [47].

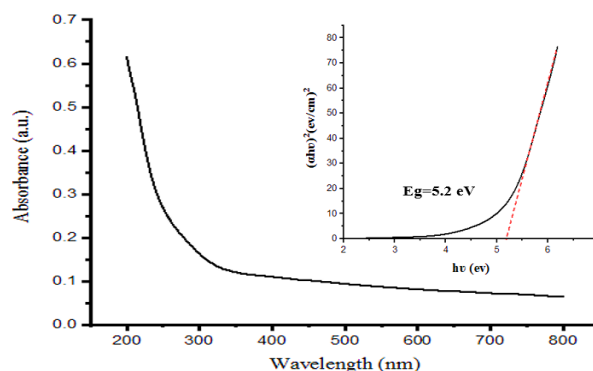


Figure 2. Absorption spectra of NiO NPs (inset: E_g of NiO by using Tauc's plot)

Table 1. Bond positions and functional groups of synthesized NiO NPs

Wavenumber (cm ⁻¹)	Bond type
3435	O=C=O Stretching
2071	O=C=O Stretching
1634	O–H Bending-Vibrational
688	Ni–O stretching-vibrational

XRD analysis of the samples' crystal structure, phase composition, and particle size makes it possible to predict and modify the antibacterial performance of the material. XRD procedure was applied to determine and verify the crystallinity structure of the produced NiO NPs. The XRD pattern of the synthesized NiO nanoparticles by the PLA method is presented in Figure 4. The XRD patterns showed no further impurity peaks. The given reflections represent a crystalline structure face-centred cubic (FCC). The XRD pattern of NiO NPs shows characteristic peaks at $2\theta = 38.4^\circ$, 44.9° , and 77.8° , corresponding to hkl values (111), (200), and (311) respectively. These diffracting peaks match the standard card number NiO (JCPDS 47-1049), which indicates nickel oxide has been created. The diffraction peaks have widened, indicating a tiny crystal size. The location and strength of these peaks, as determined by XRD analysis, reveal important details regarding the crystalline structure of the produced NiO NPs. The diffraction peak $2\theta = 52.2^\circ$ corresponds to the (102) plane representing the formation of Ni (OH)₂ (JCPDS 14-0117). For the determination of particle sizes, the Debye-Scherrer formula has been employed [48]:

$$D = (0.9\lambda) / \beta \cos \theta \quad (2)$$

In this context, D represents the crystallite size, θ is Bragg's angle, λ is the wavelength of the radiation used, and β is the full width at half maximum (FWHM) of the diffraction peak. Table 2 shows the full information of NiO NP diffraction peaks.

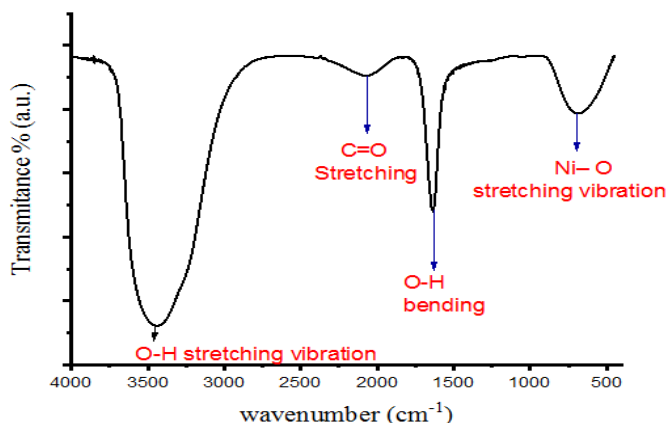
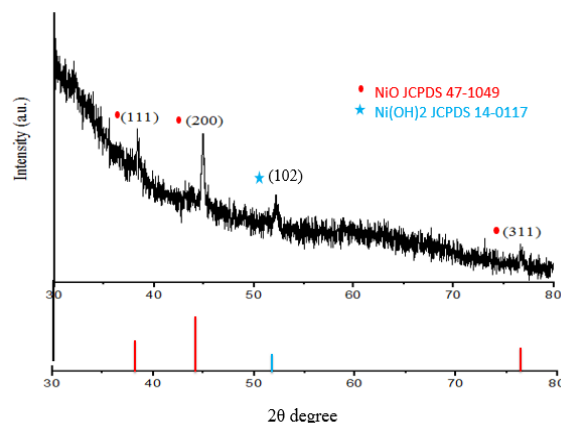
The results of the previous analysis indicate that there is a relation between the topographical surface properties of NiO NPs and their appropriateness for biological efficiency. These special characteristics of the synthesized NiO NPs can

Table 2. The structural properties of NiO NPs

2θ	(hkl)	Crystal size (Å)	D-spacing (Å)
38.4°	(111)	184	2.33971
44.9°	(200)	526	2.01509
77.8°	(311)	222	1.23952

be managed and described, allowing for the biocompatible particles to be designed for specific medical and therapeutic purposes based on how they will engage with living systems. The surface morphology of NiO NPs prepared by drying the suspended nanoparticles onto a glass substrate have been investigated using FESEM, as represented in Figure 5(a). According to several earlier investigations, MeO NP nanoparticles with sizes in the range of 1 to 100 nm and a variety of morphologies are effective antibacterial agents against infectious illnesses [49]. The NiO nanoparticles produced in this study exhibited a semispherical shape and showed agglomeration, with an average size of 44 nm, as observed in the FESEM micrograph. Thus, the nanoparticles produced in this work may represent a promising class of antibacterial agents. Image analysis, especially by software like ImageJ to measure the size distribution of NPs is well well-accepted and reliable technique in material science which is presented in Figure 5(b).

EDX is used to ascertain the fundamental basic synthesis elements of NiO NPs. Figure 6 displays the EDX spectrum for the NiO NP sample. The peaks in the spectrum of EDX between 0 and 2 keV showed that NiO NPs were synthesized effectively, according to existing findings [24, 50] since the only elements found are O, C, and Ni. The elemental analysis is displayed in the inset table of Figure 6, where the atomic percentage ratios for C, O, and Ni were 13.4, 47.5, and 39.1, respectively. These observational data collectively show that NiO NPs are prepared effectively. The elemental maps for EDX are displayed in the inset images of Figure 6. Each map is expressed in terms of colour representation of each constituent of the map as rendered in its brilliance (elementary) and black (element scarcities) maps. The elements show a uniform and consistent distribution over the whole measured region.


Figure 3. FTIR spectra of NiO NP suspension

Figure 4. XRD analysis of NiO NPs with reference cards pattern

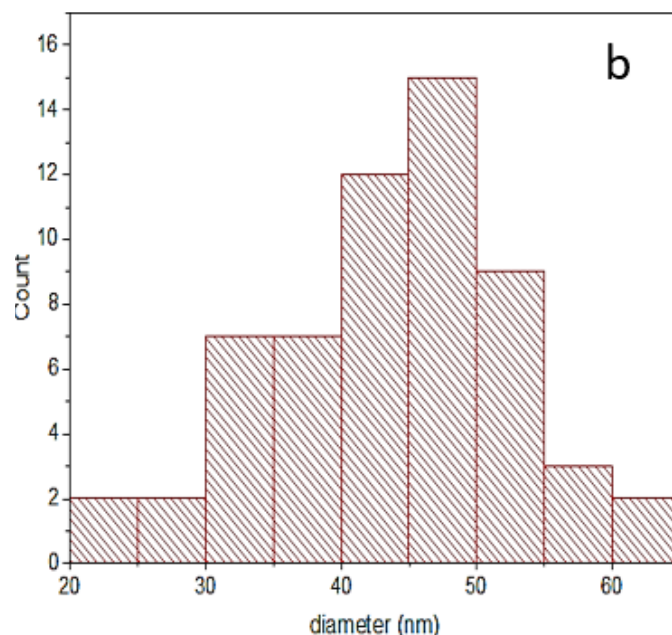
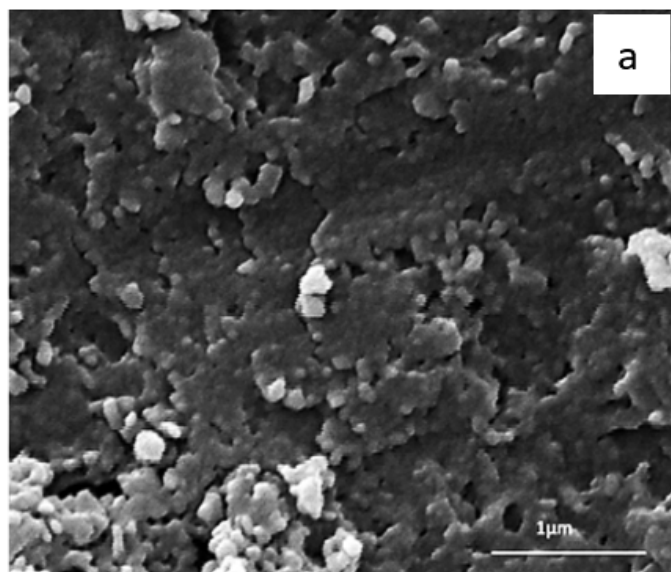


Figure 5. (a) SEM images of NiO NPs. (b) Particle size distribution

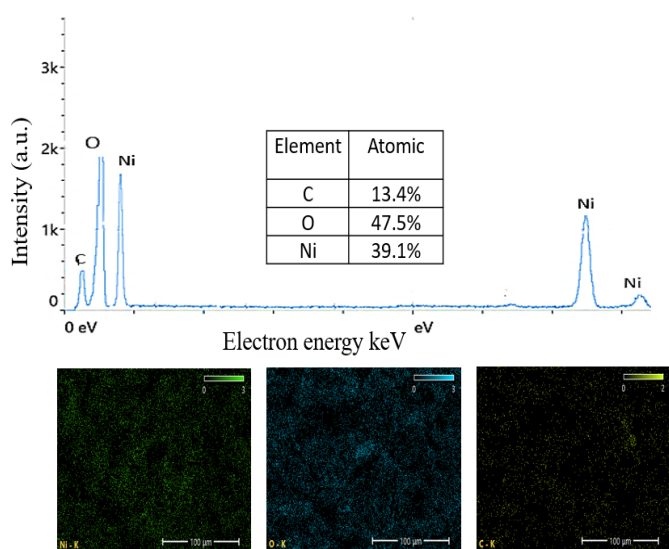


Figure 6. EDX image of NiO NPs

The zeta potential measurement of NiO NPs manufactured by PLA is shown in Figure 7.a. The zeta potential was used to estimate synthesized NiO NPs' stability in the colloidal solution. It shows how strongly particles with the same charges repel one another in the dispersion medium. Higher negative charges were found to be related to an increase in stability; that is, the greater the negative charge of NPs, the more stable they were [51]. In the current study, the negative sign of zeta potential was found (−12.5) mV. This reveals that NiO NPs produced by PLA are moderately stable. Bacterial cell walls and membranes are mostly negatively charged. The MeO NPs with positive charge and bacterial surfaces are attracted to each other due to attraction via the electrostatic interaction. Even particles with a negative zeta potential are capable of disturbing cell membranes. Therefore, interaction forces may be hydrophobic, Van der Waal, and electrostatic [52].

Figure 7(b) represents the size distribution of the NiO NPs suspension. The average particle size found for NiO NPs was 38.92 nm. It was shown that the mean size of NiO NPs obtained according to the analysis. The size ascertained by FESEM was larger than the one ascertained by DLS. The differences in the FESEM and DLS analysis conditions could account for the result situations. DLS measurements were accomplished in an aqueous medium, while FESEM analysis was done on dried samples. The drying process may influence the particle diameter determination.

The poly-dispersity index (PDI) can be assumed to be an important indicator that shows the homogeneity of the NP sizes in the sample. The PDI scale ranges from 0 to 1 [53]. A PDI value of less than (0.4) indicates that the suspension is homogeneous [54]. However, a suspension is considered exceedingly varied if the PDI value is greater than one [55]. The PDI recorded in this work was 0.189, showing that the particle size distribution of the generated NiO NPs was narrow and homogeneous. This shows that a major fraction of the NiO NPs had comparable diameters, resulting in a well-defined size distribution.

Numerous reports have been published by the research community on the application of NiO NPs in various biomedical fields. The NPs were employed as both antibacterial and antifungal agents. Thus, the cytotoxic effect and favourable biological properties of NiO NPs make these applications possible. In the present study, *S. aureus* and *E. coli* bacteria were used to examine the antibacterial behaviour of NiO NPs synthesized. The agar well diffusion assay was used to observe the growth of bacteria in culture media containing varying doses of NiO NPs by evaluating the diameter of the inhibition zone using vernier. The antibacterial properties were examined at different NiO NP concentrations (25, 50, 100, 150, and 200) μg/ml. At lower concentrations, when compared to *E. coli*, *S. aureus* demonstrated a greater sensitivity to NiO nanoparticles.

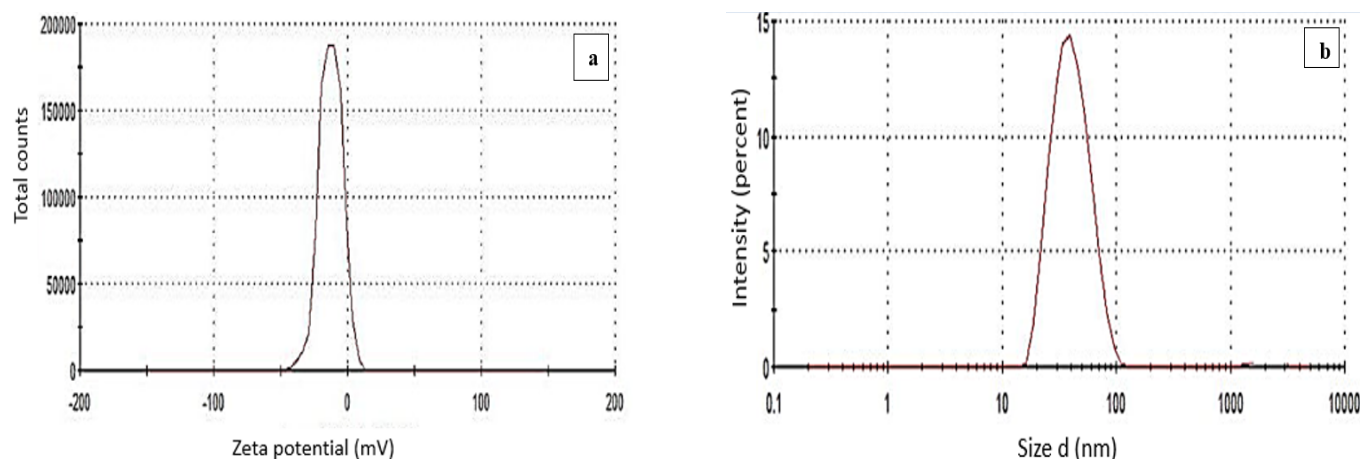


Figure 7. DLS analysis of NiO NPs (a) zeta potential, and (b) particle size

The maximum activity that appeared at 200 $\mu\text{g/ml}$ was 16 mm ($p < 0.001$). The antibacterial assay's statistical findings are displayed in Figure 8. and Figure 9 presents a photographic plate inhibition zone.

The outcomes demonstrated that NiO NPs inhibited bacterial growth in concentration-dependent manner. These results are also presented in the previous study by Jassim *et al.* [56]. Nickel oxide nanoparticles (NiO NPs) are used in the biological field due to their cytotoxic effects and beneficial biological properties. The mechanism involves the release of Ni ions when NiO NPs come into contact with bacteria; these ions then adhere to the bacterial cell membrane. This process leads to an increase in reactive oxygen species (ROS) within the bacterial cell's lipid peroxides, weakening the cell membrane. This ultimately results in cell death due to changes in the bacterial strain's membrane permeability. NiO NPs can bind to bacteria and disrupt their permeability and respiration function, depending on the large surface area available for interaction. Bacterial growth and survival are influenced by the physio-chemical characteristics of NPs, which are largely resolute according to size, shape, and concentration [57]. The minimum inhibitory (MIC) is the concentration at which, after one day of culture, a bacterium will not grow visibly. When referring to conventional pharmaceutical

antibiotics, mass concentration is used, such as $\mu\text{g/ml}$. MIC of NiO NPs was 25 $\mu\text{g/ml}$ for both strains. These findings were most widely accepted and agreed with [58, 59]. The results obtained from this work were better than previous works in terms of using a small dose of the nanomaterial to obtain the same value from the inhibition zone as reported by Vijaya Kumar *et al.* [17].

6. CONCLUSION

In conclusion, NiO NPs were successfully manufactured using PLA, as confirmed by the characterization of their optical, chemical, zeta potential, and structural properties. XRD analysis was employed to identify the structure of FCC NiO NPs. Employing FTIR analysis, the existence of every functional group was determined. The FESEM study revealed that both NiO NPs had spherically shaped morphologies. The optical absorption spectra showed the NiO NPs have a wide band gap of 5.2 eV. The synthesized NiO NPs include both Ni and O, as validated by EDX analysis. NiO NPs showed moderate zeta potential values, and the particle size was 38.9 nm. NiO NPs show remarkable antibacterial efficacy against *S. aureus* and *E. coli*, which was approximately 16 mm for both. Investigation into the antibacterial properties of NiO NPs has helped in the development of NiO NPs as effective agents versus

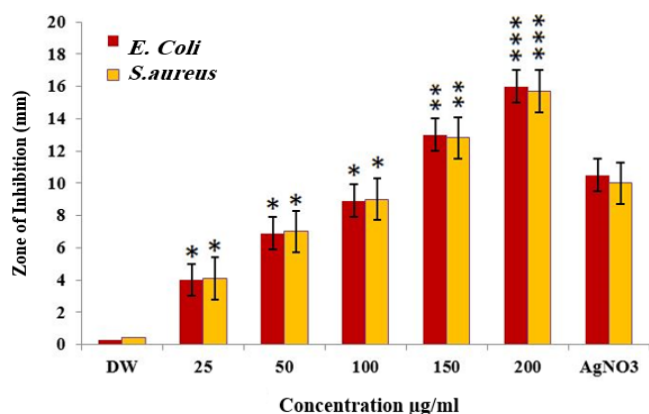


Figure 8. Antibacterial assay of NiO NPs vs. (*E. coli* and *S. aureus*). Statistics are presented as (mean \pm SD (* $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$); $n = 3$)

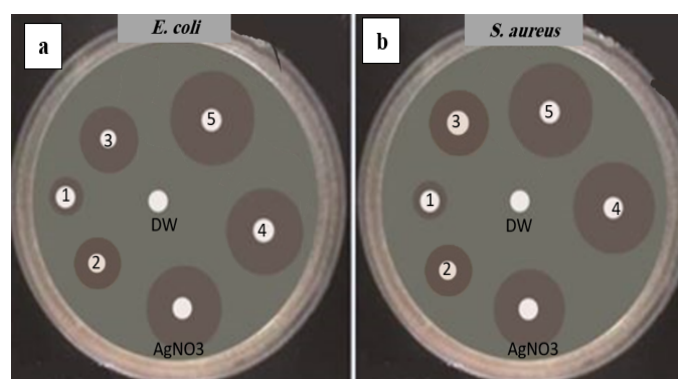


Figure 9. Zone of inhibition viewing the antibacterial test result of (a) *E. coli* (b) *S. aureus*. (1, 2, 3, 4, & 5 refer to the concentrations of NiO NPs (25, 50, 100, 150, and 200) $\mu\text{g/ml}$, respectively)

pathogenic bacteria in wounds, which highlights the promising role of NiO NPs in biomedical applications. Therefore, the obtained results emphasize the importance of NiO NPs made by a laser ablation technique in improving the overall effectiveness of antibacterial therapies and wound management strategies, and they suggest the necessity of further research and development in this field.

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